

AD_____

Award Number: DAMD17-01-1-0198

TITLE: Estrogen Receptor Inhibition of NF- (kappa)B Activity in
Breast Cancer

PRINCIPAL INVESTIGATOR: Kendall W. Nettles
Geoffrey L. Greene, Ph.D.

CONTRACTING ORGANIZATION: The University of Chicago
Chicago, Illinois 60637

REPORT DATE: June 2003

TYPE OF REPORT: Annual Summary

PREPARED FOR: U.S. Army Medical Research and Materiel Command
Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;
Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

20040830 076

REPORT DOCUMENTATION PAGEForm Approved
OMB No. 074-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing this collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503

1. AGENCY USE ONLY (Leave blank)		2. REPORT DATE June 2003	3. REPORT TYPE AND DATES COVERED Annual Summary (28 May 2002 - 27 May 2003)	
4. TITLE AND SUBTITLE Estrogen Receptor Inhibition of NF- (kappa)B Activity in Breast Cancer			5. FUNDING NUMBERS DAMD17-01-1-0198	
6. AUTHOR(S) Kendall W. Nettles Geoffrey L. Greene, Ph.D.				
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) The University of Chicago Chicago, Illinois 60637 <i>E-Mail:</i> knettles@midway.uchicago.edu			8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research and Materiel Command Fort Detrick, Maryland 21702-5012			10. SPONSORING / MONITORING AGENCY REPORT NUMBER	
11. SUPPLEMENTARY NOTES				
12a. DISTRIBUTION / AVAILABILITY STATEMENT Approved for Public Release; Distribution Unlimited				12b. DISTRIBUTION CODE
13. ABSTRACT (Maximum 200 Words) Estrogen Receptor-alpha (ER) mediated inhibition of NF-kappaB contributes to the anti-inflammatory and protective effects of estrogen in bone, cardiovascular, and breast cancer. Cross talk could be caused by direct or indirect association of these transcription factors, or by competition for other components of the transcriptional apparatus. In order to distinguish among these possibilities, we identified clonal variants of ER(+) MCF-7 breast cancer cells that either do (MCF-7 SI) or do not (MCF7 SS) display ER mediated inhibition of NF-kappaB transcriptional activity. Transient transfection of various coactivators into the MCF-7SS cells revealed that only CBP and p300 were able to promote an inhibitory effect of estradiol on NF-kappa B activity. Western blot analysis showed that CBP protein levels were reduced in this cell line relative to the MCF-7SI cells. Both immunofluorescent microscopy and co-immunoprecipitation showed an association between ER and NF-kappaB in the MCF-7SI cells. CBP also immunoprecipitated with both ER and NF-kappaB. The use of deletion constructs demonstrated that the ER ligand binding domain was necessary and sufficient for...				
14. SUBJECT TERMS No Subject Terms Provided.				15. NUMBER OF PAGES 6
				16. PRICE CODE
17. SECURITY CLASSIFICATION OF REPORT Unclassified	18. SECURITY CLASSIFICATION OF THIS PAGE Unclassified	19. SECURITY CLASSIFICATION OF ABSTRACT Unclassified	20. LIMITATION OF ABSTRACT Unlimited	

Table of Contents

Cover.....	1
SF 298.....	2
Table of Contents.....	3
Key Research Accomplishments.....	4
Statement of Work.....	6

Annual Summary Training Report for May 28, 2002– May 27, 2003

Key Accomplishments

- Defined the ER ligand binding domain as required for physical association with NF-kappaB in immunoprecipitation.
- Identified ligand binding domain of ER as necessary for trans-repression of NF-kappaB.
- Used chromatin immunoprecipitation to demonstrate that ER is recruited to the MCP-1 NF-kappaB response element in intact cells.
- Expressed and purified the ER LBD for crystallization.
- Completed subcloning of the CBP CH3 domain into a bacterial expression vector.

Training and Research Accomplishments Related to the Statement of Work (Statement of Work attached as Abstract).

Task 1: To test the hypothesis that p160 coactivators multimerize, assemble onto each LBD of an ER α dimer, and are redundant for ER α , but not for NF- κ B function.

- 1a. Construction and purification of GST-ER completed.
- 1b. RNAi against CBP was tested and proved effective in reducing CBP protein expression in cell lines.
- 1c. A series of deletion mutant ER constructs were used to identify the region of ER that interacts with NF-kappaB in an immunoprecipitation assay. These experiments clearly identified the ER ligand binding domain as necessary and sufficient for the association. A series of CBP deletion constructs were also used to identify the regions of CBP required for mediating suppressive cross-talk in a transient transfection assay. Deletion of the p160 interaction domain in the CBP c-terminus had no effect, as this construct was still able to facilitate an estradiol mediated suppression of NF-kappaB transcription in the MCF-7SS cells. Cos-7 cells were used to test for dominant negative effects of deletion CBP constructs on ER- NF-kappaB cross-talk. Deletion of the p65 interacting domain or the CBP CH3 domain allowed CBP to act in an dominant negative fashion, blocking the estradiol mediated suppression of NF-kappaB. The GST-ER-ligand binding domain was used to demonstrate a direct interaction between the ER and the CBP CH3 domain.
- 1d. The effects of CBP RNAi will be examined on NF-kappaB transcription during months 24-36.
- 1e. As detailed in last years report, this assay was replaced with a variety of other experiments designed to probe for associations.

Task 2: To test the hypothesis that coactivators binding directly to ER α and NF-kappaB mediate the inhibitory interactions between them.

- 2a. Association of ER with HSP90 was not examined at the recommendation of my thesis committee.
- 2b. Luciferase assay developed and tested
- 2c. Transient transfection assays developed and completed. These experiments identified the regions of CBP required for ER mediated suppression of NF-kappaB, namely the p65 interaction region in the n-terminus, and the ER interaction region in the CH3 domain. Notably, the p160 interaction domain in the c-terminus was not required.
- 2d. A PCAF expression plasmid was obtained for another investigator.

2e. Overexpression of PCAF blocked the suppressive effects of estradiol in transient transfection assays, suggesting the competition between ER and PCAF for the CH3 domain may be the fundamental mechanism of repression. P65 DNA binding activity was measured with DNA gel shifts and MCF7 extracts, demonstrating that there was no loss of DNA binding activity associated with estradiol treatment.

Task 3: Examine the role of the CH3 domain of CBP in the ER-mediated inhibition of NF-kappaB, and the structural basis of the CH3 ER association.

1a. These experiments will be performed during months 24-36.

1b. Binding of ER to the MCP-1 NFkappaB response element DNA sequence was demonstrated with the chromatin immunoprecipitation assay. Binding of PCAF will be examined during months 24-36.

1c. Completed

1d. Expression and purification of the ER ligand binding domain is completed. Purification of the CH3 domain will occur during onths 24-36.

1e-f. These experiments will be performed during months 24-36.

Statement of Work

Task 1: to test the hypothesis the p160 coactivators multimerize, assemble onto each LBD of an ER α dimer, and are redundant for ER α , but not NF- κ B function.

- a. Develop and purify ER and GST-ER-LBD and dimerization defective GST-ER-LBD proteins, including mutagenesis of the GST-eR-LBD plasmid (months 1-6).
- b. Develop and characterize p160 antisense nucleotides in cell culture, including northern and western blot analysis for the p160 family members. (months 1-12).
- c. Perform pull-down assays. (months 1-18).
- d. Perform transient transfection assays with antisense nucleotides (months 12-24).
- e. Confirm protein interactions with yeast two hybrid system (months 24-36).

Task 2: To test the hypothesis that coactivators binding directly to ER α and NF-kappaB mediate the inhibitory interactions between them.

- a. Purify ER from MCF-7 extracts under conditions that allow retention of chaperone complex, followed by western blot analysis for ER, p65, and HSP90 (months 1-12).
- b. Develop luciferase reporter with estrogen and NF-kappaB response elements. (months 1-6)
- c. Perform transient transfection assays with transcriptional coactivator expression plasmids (months 6-24), verifying ER and p65 protein expression by western blot.
- d. Perform transient transfection of ASC-1 and PCAF, determination of NF-kappaB and ER DNA binding activity in cell extracts (months 24-36).

Task 3: Examine the role of the CH3 domain of CBP in the ER-mediated inhibition of NF-kappaB, and the structural basis of the CH3 ER association.

- a. Test if in vitro translated PCAF competes away CH3 CBP binding to GST-ER (months 12-24).
- b. Examine if ER competed for PCAF binding to the NF-KappaB enhancer of the MCP-1 gene using chromatin immunoprecipitation (months 12-36).
- c. Clone the CH3 domain of CBP into a bacterial expression vector (months 12-14).
- d. Express and purify CH3 CBP and ER (months 12-16).
- e. Cocrystallize ER and CH3 CBP (months 16-24).
- f. Crystal structure determination (months 24-36).